

Remediation of Oil- Contaminated wetland by *Hevea brasiliensis*

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ABSTRACT

The present investigation provides data on the performance of *Hevea brasiliensis* at site clean - up. Effluent sample from a wetland around abandoned oil - well head in Mgbuoba, a Nigerian Niger Delta community was characterized and a 43d hydroponic experiment using *H. Brasiliensis* in the presence of white light and salted regime, were done to assess its performance at contaminant removal. Deionized water was used as control and all experiment was done in triplicates. Observed levels of studied heavy metals (Cd, Pb, and Fe) were statistically significant at $p \leq 0.05$ and were reduced by 98 % and 91 % respectively for Salted and Unsalted regimes. Salt treatment showed greater root uptake and were up to 58 % (Cd) and 59 % (Pb) compared to the unsalted 42 % and 41 % respectively. Generally, results indicate excellent levels of phytoextraction of Cd and Pb leaving relatively low or non – detectable values, thus presenting *H. brasiliensis* as a possible candidate for phytoremediation of oil-related contaminated liquid effluent especially a salt – assisted design.

Keywords: Oil-related effluent, Heavy metals removal, *Hevea brasiliensis*, Hydroponic experiment, Salt treatment.

INTRODUCTION

Wetlands are part of the foundation of our nation's water resources and are vital to the health of waterways and communities that are downstream. Wetlands feed downstream waters, trap floodwaters, recharge groundwater supplies, remove pollution, and provide fish and wildlife habitat. Wetlands are also economic drivers because of their key role in fishing, hunting, agriculture and recreation¹. The health of the environment and the lives of people are intertwined with the health of the water system. The food, water, and cultural identity of many local peoples are closely related to the delta ecosystem. The rivers and streams are used for drinking, bathing, fishing, harvesting, and fermenting cassava². When these water sources are polluted, it may pose health risk to the people³. Pollution is an undesirable change in the physical, chemical or biological characteristics of air, water and soil that may harmfully affect the life or create a potential health hazard for any living organism and in particular man⁴. It is well known that environmental pollution is a product of urbanization and technology, and other attendant factors of population density, industrialization and mechanization that serve to provide the necessities of the population^{5,6,7}. Anthropogenic sources of pollution include those associated with fossil fuel, coal combustion, and industrial effluents⁴. Effluents discharged into rivers, which may affect aquatic animals like fish, may do so either directly or indirectly⁸ and when an environment is polluted, it may require remediation. Remediation is generally used to mean the clean-up or making safe of a site or water body contaminated by toxic substances, whether they are natural or man-made⁹. The use of higher plants in remediation of metal contaminated sites, known as phytoremediation, is gaining worldwide importance due to its low cost involvement and eco-friendliness of the method^{10,11}. It is therefore the objectives of this study to investigate clean – up potential of *H. Brasiliensis* at removal of Cadmium and Lead from polluted wetland and the influences by salting.

MATERIALS AND METHODS

Sample sourcing

Samples were collected midstream at random with sterile two (2) litres plastic containers from the wetland and immediately corked with the lid preserved in a container of at 4⁰C and taken to the laboratory within twenty-four (24) hours for initial physico-chemical analysis while observing necessary protocols. *H. brasillensis* of about eight weeks old were sourced from an extension of the Delta Rubber Plantation, Elele and conveyed to Port Harcourt all in Rivers State, Nigeria while observing the necessary protocol.

Experimental Set-up

Initial characterization of relevant parameters (pH, temperature, conductivity, salinity, Total Dissolved Solid, Chemical Oxygen Demand, Biological Oxygen Demand, Dissolved Oxygen, Phenol, Cyanide, Cadmium, Mercury, Lead, Chromium, Iron, and Turbidity) were assayed immediately after collection of samples.

Initial measurements of leaf length, leaf breadth, leaf area, root and stem length were taken. The rubber plants were planted hydroponically so that only the roots with some lower parts of the stem were immersed in samples. The first set of plants was immersed in a beaker containing 1.2 litres of deionized water as a control. The second, containing effluent sample of equal volume as were in the first beaker while the third had in addition to the second, salt augmentation (50Mm of NaCl). All experiment was performed in triplicates and plants were exposed to florescent light at night at room temperature (43d). Illumination with florescent white light increased enzyme (peroxidase) content and the treatment with NaCl stimulated excretion of peroxidase from the cells of the plant roots¹² and this suggests that production and product recovery from plant cells can be greatly improved by application of combined treatments.

Determination of Physical Characteristics of the Sample

The pH meter (H18314-Hanna) was calibrated with phosphate buffer solutions pH determination. A micro thermometer's probe was immersed into samples and temperature measurements read off at a steady reading. In determining the chemical oxygen demand (COD), the permanganate method was employed. For each effluent sample a blank sample was prepared and 10ml of 20% H₂SO₄ was added to each sample after which, 10ml of 80N potassium permanganate was added until a pink colouration was observed. The samples and the blanks were transferred to the water bath for 30 minutes at 100⁰C. Agavn, 3mg of 10% potassium iodide and 19g starch were added to each sample and their blanks. An 80N Na₂SO₃ was titrated against each sample and its blank, and the chemical oxygen demand was calculated as given: COD= (Blank - Sample (titrant) x 10) / (Blank x Sample (titrand)).

The concentration of BOD was determined by dividing the COD by a factor of 1.5

Dissolved Oxygen (DO) / Electrical conductivity/ TDS/ Turbidity Determination: Electrometric method was employed using a DO meter/ conductivity meter (Hanna combo HI 98129) / Hach 2100AN turbidimeter respectively.

Determination of Chemical Characteristics of the Sample

Salinity: Firstly, the chloride concentration of the sample was determined. This was achieved by adding two drops of dichromate (K₂CrO₄) as an indicator to the sample and Silver nitrate (AgNO₃) was titrated against it with stirring until the slightest perceptible reddish coloration persisted. Determination of another sample (blank) was carried out and chloride concentration calculated. The salinity of the sample is the product of chloride(Cl⁻) level and the factor (1.65)

Total Suspended Solids (TSS):The UV spectrophotometer (Hach DR2400) was standardized with deionized water in the cuvette. Some of the sample was poured into a cuvette and placed in the sample compartment. The absorbance was read-off at the wavelength of 810nm.

Total Hydrocarbon (THC):Two hundred (200) milliliters of the samples was measured into a beaker and 20 ml of chloroform (CHCl₃) was added to it. The solution was homogenized and allowed to stand for 2min for self - separation. Cotton wool was inserted in a separating funnel placed in a beaker. Sodium

sulphate (dehydrating agent) and silica gel were added to the cotton wool for purity. The solution was poured into the funnel and the tap opened to release the extract into the beaker. The total hydrocarbon was gotten by the subtraction of the volume of the chloroform from that of the extract.

Phenol Determination: The concentration of phenol was determined using phenol pillow reagent and UV spectrophotometer (Hach DR2400). The absorbance of the sample was read-off at 460nm wavelength.

Cyanide Determination: The determination of cyanide was done using UV-spectrophotometer (Hach DR2400). The device was set to cyanide-determination mode and standardized. The absorbance value was read-off at 612 nm wavelength.

Heavy metals: Plant samples were first digested to reduce organic matter interference and convert metal to a form that can be analyzed by Atomic Absorption Spectrophotometry, AAS. Analyses were done employing AAS Model GDC Avanta A6600 at corresponding wavelengths using appropriate cathode lamp and results in mg/l were recorded.

Statistical Analysis: At 95 % confidence level, means of triplicate determinations were subjected to ANOVA using Microsoft Windows Excel 2007.

RESULTS AND DISCUSSION

Physicochemistry of Wetland samples

The results for the physico-chemical properties and levels of some major contaminants in water samples from a wetland around an abandoned oil - well in Mgbuoba Community for various regimes are given in table 1. The pH of the control sample was less alkaline (6.52) than salted (8.78) and unsalted (8.86) and compares favourably with WHO ¹³ set limit for drinking water. Electrical conductivity (EC) is the numerical expression of an aqueous solution to carry electrical current and is a useful indicator of the mineralization in a water sample ¹⁴. The turbidity of the sample prior to treatment (245.7 NTU) decreased in the salted regime (6.56 NTU) and the unsalted (7.36 NTU). This observed value shows that the salted regime had more effect in turbidity clearance. The biological oxygen demand (BOD) for the salted regime increased to 3.242mg/l at day 23 (values not shown) when compared to the initial (1.26mg/l) but reduced at day 43 (0.42 mg/l) and in the unsalted (0.75mg/l). The salted may have encouraged microbial activity in the sample as well as peroxidase activity ¹⁵.

Table 1: Mean levels of sample physicochemistry (before and at the end of experiment)

S/N	Parameters	End of Experiment			
		Before Experiment	control	unsalted	salted
1.	pH	8.24 ^a ± 0.004	6.52 ^b ± 0.01	8.86 ^c ± 0.01	8.78 ^c ± 0.004
2.	Temp(°C)	25.6 ^a ± 0.06	26.5 ^b ± 0.1	27.9 ^c ± 0.06	28.5 ^d ± 0.07
3.	BOD(mg/l)	1.26 ^a ± 0.002	0.02 ^b ± 0.006	0.745 ^c ± 0.001	0.42 ^c ± 0.002
4.	DO (mg/l)	3.37 ^a ± 0.002	5.6 ^b ± 0.007	4.90 ^c ± 0.03	5.20 ^d ± 0.03
5.	COD (mg/l)	1.88 ^a ± 0.006	0.03 ^b ± 0.002	1.12 ^c ± 0.001	1.27 ^d ± 0.006
6.	Conductivity(µS/cm)	106 ^a ± 1.52	226 ^b ± 1	187 ^c ± 1.15	14320 ^d ± 10.8
7.	Turbidity (NTU)	245.7 ^a ± 0.3	0.93 ^b ± 0.003	7.36 ^c ± 0.03	6.56 ^d ± 0.007
8.	Salinity (mg/l)	16.5 ^a ± 0.21	16.5 ^a ± 0.06	30 ^b ± 1.15	4000 ^c ± 20.61
9.	THC (mg/l)	10.27 ^a ± 0.006	0.02 ^b ± 0.0002	2.05 ^c ± 0.02	6.91 ^d ± 0.01
10.	TSS (mg/l)	510 ^a ± 4.6	2.0 ^b ± 0	8.0 ^c ± 0.15	9.0 ^d ± 0

11.	TDS (mg/l)	54.0 ± 0.91	115.0 ± 1.22	95.0 ± 0.50	7520.0 ± 18.71
12.	Phenol (mg/l)	0.15	0.00	<0.001	<0.001
13.	Cyanide (mg/l)	<0.001	0.00	<0.001	<0.001
14.	Cd (mg/l)	0.082 ± 0.002	0.00	<0.001	<0.001
15.	Hg (mg/l)	<0.001	0.00	<0.001	<0.001
16.	Pb (mg/l)	0.374 ± 0.006	0.00	<0.001	<0.001
17.	Cr ⁺ (mg/l)	<0.001	0.00	<0.001	<0.001
18.	Fe (mg/l)	24.55 ± 0.003	0.00	0.31 ± 0.001	0.59

Values are means ± SEM; Means in the same row with same alphabets are not significantly different at $P \leq 0.05$

Elevated levels of Cadmium, Lead and Iron were observed alongside Total hydrocarbons in Wetland samples. These levels were however, reduced to non – detectable or tolerable levels at the end of the experiment. Also, statistically significant decreases in COD and BOD were observed after 43d and did not conform with given Department of Petroleum Resources guideline. On the contrary, significant increases were observed for DO and that is attributable to increased removal of organics and suspended solids as shown in table 1. Respiration, activity and growth rate of fresh water organisms generally increase as temperature and dissolved oxygen increase up to a threshold¹⁶. Improved levels of dissolved oxygen were more marked in the salted regime (5.20mg/l) than in the unsalted (4.90mg/l), thus, suggesting that the salted regime could be more tolerable to aquatic lives.

Increased salinity, conductivity and salinity values for salt – treated regime at 43d could be attributed to salt treatment. Efficient Phenol removal was also observed for all treatments. Cyanide, Mercury and Chromium were not detected in any of the samples. The advantages of this local plant species reported by Malik and Biswas¹⁰ as a remedial measure is markedly interesting.

Uptake expression by *H. brasillensis*

It is worthy of mention that plant tissue exhibited great resistance to posed phytotoxicity by all indices employed and it is typical of toxicity described by Pourrut et al.¹⁷ and Nafiseh et al.¹⁸. Mean contaminants uptake by *H. Brasillensis* is as given in table 2. Tissue analysis of root materials indicated evidence of some heavy metals uptake as shown in table 2. The higher load of Cd and Pb occurred in the roots tissues of plants in the salted regimes than in the unsalted. The study shows that salt (NaCl) induced significant positive effect in the uptake of the Cd (up to 0.05mg/kg), Pb (up to 1.82mg/kg) when compared to the unsalted; (0.06mg/kg and 0.07mg/kg) respectively.

Table 2: Mean uptake of some contaminants (mg / kg) in the root section of the three regimes

S/N	Parameters	Control Sample	Unsalted Sample	Salted Sample
1.	Cd	0.00	0.06a±0.001	0.05b±0.007
2.	Hg	0.00	<0.001	<0.001
3.	Pb	0.00	0.07a±0.01	0.18b±0.01
4.	Cr	0.00	0.02a±0.005	0.03b±0.01
5.	Fe	20.30a	74.8b±1.35	58.9c±1.59
6.	Phenol	0.00	0.03a	0.11b

Observed levels of Chromium in root tissues of *H. Brasilensis* may have been introduced from nursery as levels were below detectable limits for the wetland samples. However Fe concentration in the root of the salted (58.9mg/kg) was less compared to the unsalted (74.8mg/kg) and that suggests deposition of Fe in other parts of the plant like the stem and leaves and this has ecological implications¹⁹. Significant phenol removal by *H. Brasilensis* is comparable to the findings of Meikap and Rot²⁰.

CONCLUSION

Generally, results indicate excellent levels of phytoextraction of Cadmium, Lead, Iron and Phenol leaving relatively low values in water sample especially with salting. Following obtained results, tissue analysis should be extended to the stem and leaves for a better understanding of contaminants distribution. Further research should be done comparing removal of such contaminants by *H. brasillensis* with other tested and confirmed plants.

REFERENCES

1. USEPA. US Environmental Protection agency. Water: Wetlands. <http://water.epa.gov/type/wetlands/index.cfm>. Accessed (2013)
2. Amnesty International Nigeria: Petroleum, Pollution and Poverty in the Niger Delta. Amnesty International Publication. Amnesty International Secretariat, Peter Benenson House 1 Easton Street London WC1X 0DW. 14 – 143 (2009)
3. Yilmaz, F., Demirak, A., Tuna, A.T. and Ozdemir, N. Heavy metals in water, sediment and tissues of *Leuciscuscephalus* from a stream in southwestern Turkey. *Chemosphere* **63**: 1451–1458 (2006)
4. Flower, L. Environmental Pollution -Especially Air Pollution - and Public Health. *Assumption University Journal of Technology*. **10 (1)**: 29-37 (2006)
5. Olade, M. A. Heavy Metal Pollution and the Need for Monitoring: Illustrated for Developing Countries in West Africa. Lead, Mercury, Cadmium and Arsenic in the Environment. Edited by T. C. Hutchinson and K. M. Meema. Published by John Wiley & Sons Ltd. 335-341 (1987)
6. Reza, R., and Singh, G. Heavy metal contamination and its indexing approach for river water. *International Journal of Environmental Science and Technology*. **7 (4)**: 785-792 (2010)
7. Sekabira, K., Origa, H. O., Basamba, T. A., Mutumba, G. and Kakudidi, E. Heavy metal assessment and water quality values in urban stream and rain water. *International Journal of Environmental Science and Technology*. **7 (4)**: 759-770 (2010)
8. Asia, I. O., Ekpo, K. E., Amayo, K. O. and Jegede, D. A. Determination of lead, cadmium and mercury in surrounding water and organs of some species of fish from Ikpoba river in Benin city, Nigeria. *International Journal of Physical Sciences*. **3 (11)**: 289-292 (2008)
9. CRC CAREFact Sheet 3 & 4 Remediation/ Bioremediation. Retrieved online from www.crccare.com on (2007)
10. Malik, N. and Biswas, A. K. Role of Higher Plants in Remediation of Metal Contaminated Sites. *Scientific Reviews & Chemical Communications*. **2 (2)**: 141-146 (2012)
11. Nwaichi, E. O., Osuji, L. C. and Onyeike, E. N. Evaluation and decontamination of crude oil-polluted soils using *Centrosema pubescens* Benth and amendment-support options. *Int. J. Phytoremediation*. **13 (4)**: 373-382 (2010)
12. Kimura, T., Kato, Y., Uozumi, N., Honda, H. and Kobayashi, T. Peroxidase production and excretion from horseradish hairy roots by light, NaCl and Peroxidase Adsorption in Situ. *Plant Tissue Culture Letters*. **8 (3)**: 158-165 (1991)
13. WHO Water for Health. Guidelines For Drinking – water Quality. Incorporating First and Second Addendum to Third Edition. Recommendations. World Health Organization, Geneva, Switzerland **1**: (2010)

14. Jain, S.K., Awasthi, A.M., Jain, N.K. and Agrawal, G.P. Calcium silicatebased microballoons of repaglinide for gastro-retentive floating drugdelivery: Preparation andin-vitrocharacterization. *Journal of Controlled Release*.**107**: 300-309 (2005)
15. Geiger, J. P., Rio, B., Nandris, D. and Nicole, M. Peroxidase production in tissues of the rubber tree following infection by root rot fungi. *Physiological and Molecular Plant Pathology*. **34 (3)**: 241-256 (1989)
16. Harding, J. S., Moore, T. A., Black, A., Centeno, J. A. and Trumm, D. A. Impacts of metals and mining on stream communities: Metal Contaminants in New Zealand, resolutionz press, Christchurch,NZ. 343-357 (2005)
17. Pourrut, B., Shahid, M., Dumat, C., Winterton, P. and Pinelli, E. Lead Uptake, Toxicity, and Detoxification in Plants. *Reviews of Environmental Contamination and Toxicology*. **213**: 113-136 (2011)
18. Nafiseh, N., Mehrdad, L. and Ali, G. Accumulation of chromium and its effect on growth of (*Allium cepa* cv. Hybrid). *European Journal of Experimental Biology*. **2 (4)**: 969-974 (2012)
19. Gulfraz, M., Mussaddeq, Y., Khanum, R. & Ahmad, T. Metal contamination in Wheat crops (*Triticum estivum* L.) irrigated with industrial effluents. *J. of Biological Sci.* **3(3)**: 335-339 (2003)
20. Meikap, B. C. and Rot, G. K. Removal of Phenolic Compounds from IndustrialWaste Water by Semifluidized Bed Bio-Reactor. *Journal of the IPHE*. **3**: 54-61 (1997)