

Morphotypes of Mycorrhizal Fungi of Cymbidium Species

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ABSTRACT

Mycorrhizal association is known to be important to orchid species and a complete understanding of the fungi that form mycorrhizas is required for orchid ecology and conservation. Cymbidium is a orchid found in Eastern and Western Ghats of India at high altitudes. Previously, we found the genetic diversity of this species has been reduced recent years due to habitat destruction and fragmentation, but little was known about the relationship between this orchid species and the mycorrhizal fungi. The Rhizoctonia-like fungi are the commonly accepted mycorrhizal fungi associated with orchids. In this study, morphotypes of the Rhizoctonia-like fungi associated with Cymbidium species were investigated and their isolates were obtained.

Key words: endophytes, Cymbidium species, Morphotypes and Rhizoctonia-like fungi.

INTRODUCTION

The mycorrhizal association is ubiquitous but very important symbiosis in nature, which plays an essential role in the maintenance of most terrestrial ecosystems¹. Over 90% of all plant species can form mycorrhizas with different kinds of fungi and the existence of mycorrhizal fungi can confer to their hosts many adaptive advantages via improved water and nutrient or minerals uptake from the soil^{2,3,4}, enhanced plant growth^{5,6}, reduced toxic element accumulation^{1,7} and increased resistance to pathogen damage⁸. The Orchidaceae, which is one of the largest and

most diverse plant families, is distributed worldwide⁹. However, many orchid species have suffered dramatic declines in distribution and some species have become rare and endangered in recent decades¹⁰. Mycorrhizal association is known to be important to orchids because they depend on the presence of suitable fungal partners for seed germination and seedling development^{11,12}. Therefore, a complete understanding of the mycorrhizal fungi of many threatened orchid species is required for conservation action plans¹³.

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The study of the earliest diverging orchid lineages and distribution of mycorrhizal fungal associates across orchid phylogeny supported that the ancestral state is an association to the Rhizoctonia-like fungi lineages¹⁴ and orchid mycorrhizas are predominantly represented by associations between photosynthetic plants and Rhizoctonia-like fungi¹³. The Rhizoctonia-like fungi includes members of the Ceratobasidiaceae, Sebaciniales and Tulasnellaceae^{13,14}. Many fungi that have been isolated from orchid roots have been identified as Rhizoctonia-like. Members of that group do not form asexual spores and all share certain distinctive vegetative characters. Six Epulorhiza species have been described based on the shape and the dimensions of monilioid cells¹⁵.

Cymbidium genus is distributed in tropical and subtropical Asia (such as northern India, China, Japan, Malaysia, the Philippines, and Borneo) and northern Australia. The larger flowered species from which the large flowered hybrids are derived grow at high

altitudes¹⁶. Several species of Cymbidium occurring in Eastern North Asia, e.g., Cymbidium goeringii and C. kanran, are important in oriental floricultural industries. However, these could not properly performed without any OM fungi. Recently, the inoculation of OM fungal isolates to Cymbidium hybrids was reported to increase survival rates, and they protected plants from root rots or soil pathogens¹⁷. The present work involves isolation and identification of mycorrhiza of *Vanda* and further studies can be done by molecular methods.

MATERIALS AND METHODS

Study area and Location

Roots were procured from three different regions of naturally growing plants of Cymbidium that is shown in the table.1. These were collected in October to January during their active vegetative growth, stored in the paper bags/ziplock and transferred to the laboratory.

Table 1. Location of the different populations of *Cymbidium* species

Population Code	Location
PU14	Kodai
KEKU	Kulitholu, Kerala
THO	Thodu hills, Kerala

Isolation of fungal endophytes

Potential mycorrhizal fungi were isolated from the orchid plants and identified from pure culture of *Cymbidium species*, unlike many other temperate, terrestrial orchids which have thick roots and often produce abundant pelotons, has few active pelotons suitable for isolation. Thus, endophytic fungi were isolated from single hyphal tips emerging from sterilized root portions as in the isolation method described¹⁸. Three roots per plant were carefully cleaned from the soil under running water, surface-sterilized in 0.1% (v/v) Mercuric chloride and 75% (v/v) ethanol for 3 min and 5 s respectively, and subsequently

washed three times in sterile distilled water. Root sections of 3–5 mm thickness were obtained by cutting and placed in a petridish with potato dextrose agar (PDA). Petri dishes were incubated at 25°C in the dark and observed for fungi growing every 2 days for at least 3 weeks. The growing colonies were separated onto fresh media for purity and this process was repeated three times.

RESULTS AND DISCUSSION

Morphological identification

Classification of the endophytic fungi was based on their growth rate and morphological characteristics, including colonial morphology,

production of conidiogenous cells, conidial size and dimension on PDA medium¹⁹ and similar isolates were grouped into one morphotype. The Rhizoctonia-like fungal endophytes were recognized by the following characteristics: hyphas hyaline with

constricted branch points, 2.5–9 μ m diameter; submerged growth in PDA; ellipsoid, globose or irregular monilioid cells; colony creamy white to pale tan or orange, rubbery or leathery in appearance and texture^{20,21,22}.

Table 2. Morphotypes of fungal isolates from the *Cymbidium* species

Morphotype	Colony color	Colony texture	Growth rate (cm/day)	Aerial mycelium	Conidial shape	Conidia size
I	Creamy white	Bushy	3.25	Absent	Circular	-
II	Brown to white	Leathery	6.5	Absent	Irregular, cylindrical	-
III	Greenish brown to white	Brushy, Slightly loose	6.2	Absent	Elliptic irregular	-

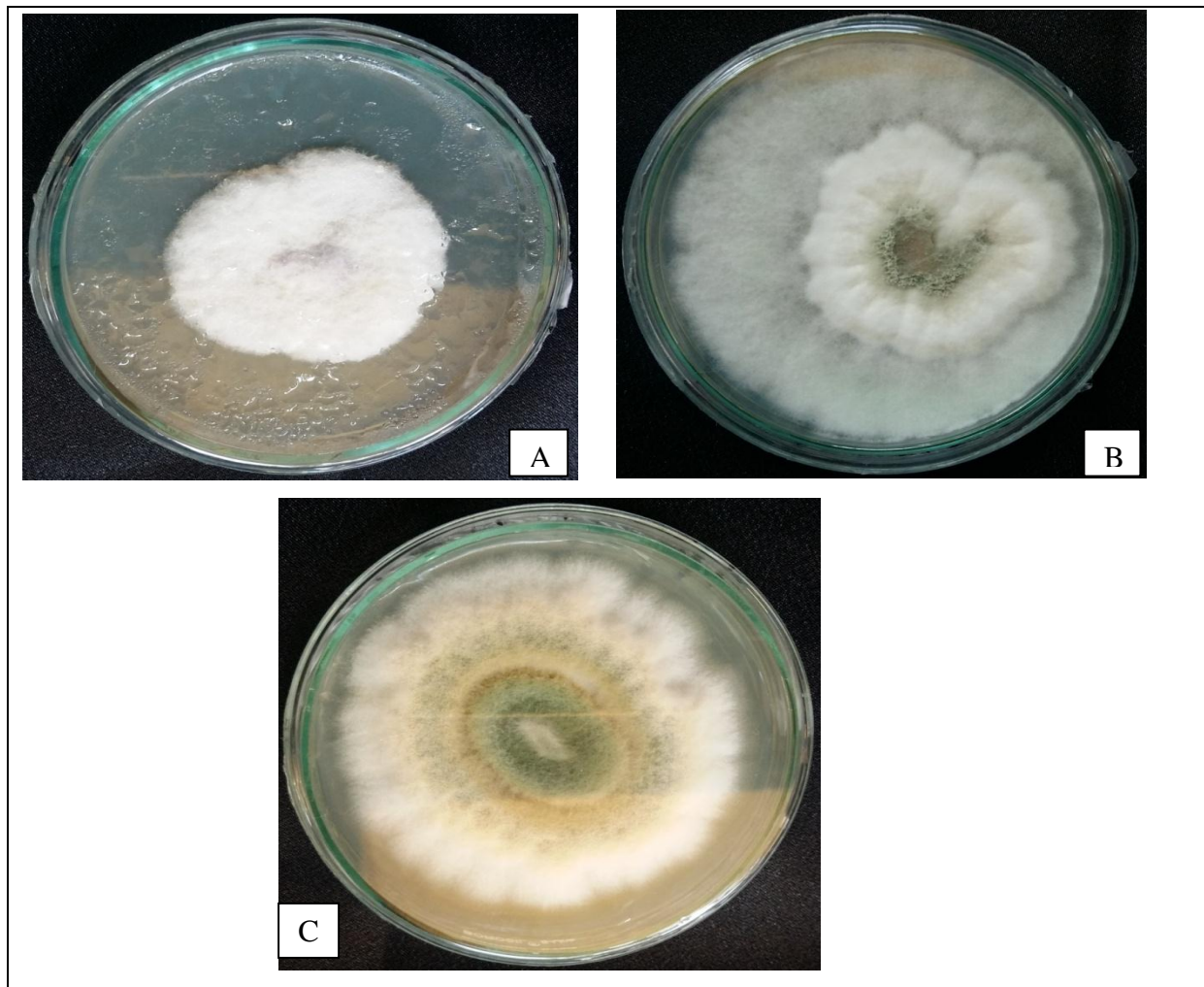


Fig. 1: The morphotypes of endophytes isolated from *Cymbidium* species

After isolation and purification, isolates of endophytic fungi from the plants were obtained. According to their morphological characters and growth rate on PDA medium, fungal isolates were classified into different morphotypes (Table 2). Morphological characters and detailed descriptions of the morphotypes were given in Fig. 1 and Table 2. Only the Rhizoctonia-like isolates were taken and subjected to further studies for DNA extraction and phylogenetic analysis.

CONCLUSION

Mycorrhizal association is known to be important to orchid species and a complete understanding of the fungi that form mycorrhizas is required for orchid ecology and conservation. Thus, the morphotypes of the Rhizoctonia-like fungi associated with *Cymbidium* species were obtained and further conformations can be done through molecular methods.

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REFERENCES

- Smith, S.E. and Read, D.J., Mycorrhizal Symbiosis. New York: Academic Press. 19–41 (2008).
- Gardes, M., An orchid-fungus marriage: physical promiscuity, conflict and cheating. *New Phytol.*, **154**: 4–7 (2002).
- Allen, M.F., Mycorrhizal fungi: highways for water and nutrients in arid soils. *Vadose Zone J*, **6**: 291–297 (2007).
- Leigh, J., Hodge, A., and Fitter, A.H., Arbuscular mycorrhizal fungi can transfer substantial amounts of nitrogen to their host plant from organic material. *New Phytol.*, **181**: 199–207 (2009).
- Artursson, V., Finlay, R.D. and Jansson, J.K., Interactions between arbuscular mycorrhizal fungi and bacteria and their potential for stimulating plant growth. *Environ Microbiol.*, **8**: 1–10 (2006).
- Porrás-Soriano, A., Soriano-Martín, M.L., Porrás-Piedra, A. and Azcón, R., Arbuscular mycorrhizal fungi increased growth, nutrient uptake and tolerance to salinity in olive trees under nursery conditions. *J Plant Physiol.*, **166**: 1350–1359 (2009).
- Fomina, M.A., Alexander, I.J., Colpaert, J.V. and Gadd, G.M., Solubilization of toxic metal minerals and metal tolerance of mycorrhizal fungi. *Soil Biol Biochem.*, **37**: 851–866 (2005).
- Yao, M.K., Tweddell, R.J. and De Silets, H., Effect of two vesicular-arbuscular mycorrhizal fungi on the growth of micropropagated potato plantlets and on the extent of disease caused by *Rhizoctonia solani*. *Mycorrhiza*, **12**: 235–242 (2002).
- Dressler, R.L., Phylogeny and classification of the orchid family. Portland: Dioscorides Press (1993).
- Swarts, N.D. and Dixon, K.W., Perspectives on orchid conservation in botanic gardens. *Trends Plant Sci.*, **14**: 590–598 (2009).
- Bidartondo, M.I. and Read, D.J., Fungal specificity bottlenecks during orchid germination and development. *Mol Ecol.*, **17**: 3707–3716 (2008).
- Rasmussen, H.N., Recent developments in the study of orchid mycorrhiza. *Plant Soil* **244**: 149–163 (2002).
- Dearnaley, J.D.W., Martos, F. and Selosse, M.A., Orchid mycorrhizas: molecular ecology, physiology, evolution and conservation aspects. In: Hock B, editors. *Fungal Association*, 2nd edn. Berlin: Springer-Verlag. 207–230 (2012).
- Yukawa, T., Ogura-Tsujita, Y., Shefferson, R.P. and Yokoyama, J., Mycorrhizal diversity in *Apostasia* (Orchidaceae) indicates the origin and evolution of orchid mycorrhiza. *Am J Bot.*, **96**: 1997–2009 (2009).
- Ricardo, E.N., Cassio van den Berg, Olinto Liparini Pereira and Maria Catarina, M.K., Isolation and molecular characterization of *Rhizoctonia*-like fungi

- associated with orchid roots in the *Quadrilátero Ferrífero* and *Zona da Mata* regions of the state of Minas Gerais, Brazil. *Acta Botanica Brasílica.*, **28(2)**: 298-300 (2014).
16. Ji Won Hong, Hyoungmin Suh, Oh Hong Kim and Nam Sook Lee. Molecular Identification of Mycorrhizae of *Cymbidium kanran* (Orchidaceae) on Jeju Island, *Korea Mycobiology*, **43(4)**: 475–480 (2015).
 17. Lee, S.S., Lee, J.G., Lee, J.W., Park, S.Y. and Paek, K.Y., Effects of the Orchid Symbiotic fungus on the cultivations of *Cymbidium* Hybrids under the conditions of Greenhouses. *Journal of Korean Society for Horticultural Science*, **42**: 223-226 (2001).
 18. Curtis, J.T., The relation of specificity of orchid mycorrhizal fungi to the problem of symbiosis. *Am J Bot.*, **26**: 390–399 (1939).
 19. Wei, Y.K., Gao, Y.B., Zhang, X., Su, D. and Wang, Y.H. *et al.*, Distribution and diversity of *Epichloe*/*Neotyphodium* fungal endophytes from different populations of *Achnatherum sibiricum* (Poaceae) in the Inner Mongolia Steppe, China. *Fungal Divers*, **24**: 329–345 (2007).
 20. Currah, R.S. and Sherburne, R., Septal ultrastructure of some fungal endophytes from boreal orchid mycorrhizas. *Mycol Res.*, **96**: 583–587 (1992).
 21. Masuhara, G. and Katsuya, K., In situ and in vitro specificity between *Rhizoctonia* spp. and *Spiranthes-sinensis* (Persoon) Ames var. *amoena* (M. Bieberstein) Hara (Orchidaceae). *New Phytol.*, **127**: 711–718 (1994).
 22. Ma, M., Tan, T.K. and Wong, S.M., Identification and molecular phylogeny of *Epulorhiza* isolates from tropical orchids. *Mycol Res.*, **107**: 1041–1049 (2003).