

## Insilico Strategy of Epitope Design in *Clostridium botulinum*

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### ABSTRACT

*Clostridium botulinum* is the name of a collection of bacteria that frequently habitat in soil. These are rod-shaped organisms that rose under anaerobic condition. They have the ability to produce spores for their survival under unfavorable conditions. The *C. botulinum* genome comprises of 3,886,996 BP, 28.24, GC%, 3650 coding sequence, 9rRNA and 80tRNA. Identifying a legend in a drug discovery project assumes that the drug target is known and has been characterized. The majority of available drugs have protein molecules as their targets for botulinum toxins are available for botulinum toxins. In our current project, potential epitope were identified in *C. botulinum* genome (An epitope is the part of the antigen that binds to a specific antigen receptor on the surface of a B cell) the predicted epitope by using available bioinformatics tools i.e. Vaxijen, TMHMM, Bcpred, MHCpred, Propred I, Propred, T-epitope designer, phyre and Pepitope. The two screening steps were adopted in this work. Previously identified 22 essential membrane proteins sequence were taken and in the first step screening the Vaxijen, TMHMM and Bcpred were used for selecting *C. botulinum* proteins as antigenic exo-membrane B-epitope. The two epitope, one from each protein have been designed in such a way that each epitope is highly likely to bind determined the number of (HLA molecules comprising of both the MHC-I and II) and interacts with most frequent HLA alleles (A\*0201, A\*0204, B\*2705, DRB1\*0101, and DRB1\*0401) in human population. Therefore, our selected epitope are highly potential to induce both the B-cell and T-cell mediated immune responses. Of course, these selected epitope require further experimental validation.

**Key words:** insilico strategy, epitope prediction, *Clostridium botulinum*.

### INTRODUCTION

*Clostridium botulinum* is the name of a collection of bacteria that normally habitats in soil. These are rod-shaped organisms that rose under anaerobic condition. They have the ability to produce spores for their survival under unfavorable conditions. *C. botulinum* produces a toxin known as botulinum neurotoxin toxin (BoNTs) that is associated

with life threatening neuro-paralysis. BoNTs are grouped into seven different sub-types designated with alphabetic letters from A to G. Among them A, B, E and F are pathogenic to humans (CDC, 2015). This toxin is among the list of bioterrorist agent with highest potential risk to the masses and has been used as biowarfare weapon by US military since Second World War.

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The most effective treatment against BoNTs is vaccinations. Since 1940s efforts has been directed for the development of effective vaccines against botulism<sup>10</sup>. To date, under the supervision of Investigational New Drugs (INDs) a monovalent and a pentavalent vaccine is available against serotype F and A-E respectively<sup>1</sup>. While various DNA-recombinant vaccines are under clinical trial<sup>3</sup>. But, at present, there are no approved vaccines are available against<sup>4</sup>. Many different approaches are under consideration for the improved and effective diagnosis and development of treatment including vaccination, synthetic small peptides and small-molecule inhibitors<sup>11</sup>. The most effective approach for the prevention of botulism could be the vaccines consisting of suitable antigen that can elicit appropriate immune response. The process of development of effective vaccine involves recognition and identification of epitope that can stimulate strong protective humeral response<sup>17</sup>.

The pathogenicity of a pathogen is often due to its pathogenic secretory and surface proteins that have also antigenic properties too<sup>14</sup>. Therefore, can be considered as one of the potential candidates against vaccine development. B-cell generates antibodies against antigenic and pathogenic proteins after recognizing the B-cell epitope over these proteins. B-cell epitope mapping can not only beneficial for effective diagnostic procedures but can also serve as a initial step towards vaccine designing<sup>2</sup>. Likewise, the specificity and diversity between antigen and human leukocyte antigen (HLA) alleles binding also contributes to specific and selective immune response.

The epitope and peptide based vaccine developmental procedures are easy, specific and harmless as compared to orthodox

procedures. The peptide based vaccine developmental processes has accelerated as a result bacterial and human genome projects. The effective prediction and mapping of B-cells and T-cells from pathogenic bacteria with the aid of computational and bioinformatics tools is a significant achievement in the course of effective vaccine design<sup>17</sup>.

Various membrane proteins are reported in literature through bioinformatics based approach against *C. botulinum* for potential vaccine targets<sup>14</sup>. In this study, we have explored 22 different essential membrane associated proteins for peptide based vaccine design with the aid of insilco approach together with simulation and verification of fold levels. This will lead to the identification of best possible and potential epitope for both B-cell and T-cell based immunity<sup>14</sup>.

## MATERIAL AND METHODS

**Antigenic B-epitope selection:** The twenty two essential membrane proteins of *C. botulinum* were earlier identified by using of subtractive genomics approach and used as best vaccines candidates were selected for the current study and a novel approach of epitope designing was adopted where an epitope should produce both B-cell and T-cell mediated immune response<sup>5</sup>.

**Prediction of B-cell epitope:** The complete amino acids sequence of each protein was retrieved from NCBI database and all proteins sequence were analyzed at Vaxijen server for the prediction of antigenic proteins<sup>9</sup>. And default parameters with threshold value (>0.4, ACC output) were used against bacterial species to check the antigenicity of each full length protein. Proteins having antigenic score >0.5 were selected<sup>9</sup>. Then each full length proteins were subjected at TMHMM v 0.2 prediction server in order to identify the

surface exposed amino acids for each full length protein<sup>12</sup>.

In the processes of B-cell epitope prediction each full length protein was subjected at Bcpred server to Use for the identification and characterization of B-cell epitope to make a novel vaccines against *C. botulinum*. And all predicted B-cell epitope (20 mers) having a Bcpred cutoff score >0.8 were selected<sup>15</sup>. The selected were then subsequently checked for membrane topology by comparing with TMHMM results for exo-membrane amino acids sequences. Then selected B-cell was further analyzed at Vaxijen to check the antigenicity. Then finally 44 were selected for use in T-epitope selection.

**Screening one step:** The T- cell was anticipated from B-cell. Then two screening steps were approved for sequence based and QSAR simulation approaches were used in the interpretation of T- selection. In the first screening step the considerations were. The first criteria was sequence should bind with MHC class I, and class II and sequence must be >15 was selected and second sequence must bind with HLA-DRB1\*0101 and must be antigenic based on Vaxijen score. Then after this the sequence was bind with ProPred I (47 MHC Class-I alleles)<sup>15</sup>, and proPred 51 MHC Class II alleles<sup>16</sup>. And both servers were used for the determination of alleles bind with above criteria. The QSAR simulation approach was used for check the inhibitor concentration of antigenic epitope at the platform of ProPred I, ProPred and MHCpred server (DRB1\*0101) and Vaxijen, respectively<sup>6</sup>. With highest antigenicity and those bind more than 15 MHC molecules including with class I and II and less than 100nm (IC50) scores were selected.

**Screening step two:** The second screening step was subjected by using T-epitope designer

tool in which the principles was >1000 HLA fragments, peptides should bind with 75% of total HLA molecules and must bind with high score to A\*0201, A0204 and B\*2705 and must bind with DRB1\*0101, and DRB1\*0401 and MHCpred .The final list of was made with non-overlapping peptide sequences that pass these above mentioned criteria and Vaxijen and IC50 scores was used for selection of DRB1\*0101 and DRB1\*0401 binding peptides were selected for design epitope for fold level topology examination<sup>7</sup>.

#### **Homology modeling for epitope analysis:**

For the homology modeling of each protein as a whole was subjected to Phyre version 2.0 Web-server<sup>8</sup>. Followed by best model prediction on the basis of super families and e-value of template. The exo-membrane topologies of the predicted were examined through their 3D structures and clustering of with the aid of Pepitope server<sup>13</sup>. This sever utilizes 3D structure of Phyre server for the prediction of linear alignment of with their respective proteins and also for prediction their epitope pockets and exo-membrane position in 3D structure of proteins.

## **RESULTS AND DISCUSSION**

### **Antigenicity and topology of selected protein:**

By using Vaxijen the anticipated possible proteins having antigenic score >0.4 were selected as an antigenic determinant and lastly 17 proteins sequence were selected as antigenic determinants shown in **(table 1)**. The basic principle of a decent epitope is that it must be showing to cell outside and TMHMM result showed that all 17 probable antigen having surface exposed peptides shown in **(table 2)**.

<b>Table1: VexiJen predicted probable antigen and non-probable antigen</b>			
<b>S. No.</b>	<b>Protein id</b>	<b>VaxiJen score</b>	<b>Comments</b>
1	YP_001386116.1	<b>0.6282</b>	Probable ANTIGEN
2	YP_001386895.1	0.3078	<b>Probable NON-ANTIGEN</b>
3	YP_001387255.1	<b>0.4396</b>	Probable ANTIGEN
4	YP_001386503.1	<b>0.4003</b>	Probable ANTIGEN
5	YP_001386587.1	<b>0.4173</b>	Probable ANTIGEN
6	YP_001386886.1	<b>0.4931</b>	Probable ANTIGEN
7	YP_001387027.1	<b>0.7008</b>	Probable ANTIGEN
8	YP_001387384.1	<b>0.5959</b>	Probable ANTIGEN
9	YP_001387457.1	<b>0.4304</b>	Probable ANTIGEN
10	YP_001387707.1	<b>0.5763</b>	Probable ANTIGEN
11	YP_001387745.1	0.3927	<b>Probable NON-ANTIGEN</b>
12	YP_001388265.1	<b>0.5384</b>	Probable ANTIGEN
13	YP_001389313.1	0.2271	<b>Probable NON-ANTIGEN</b>
14	YP_001387020.1	<b>0.5164</b>	Probable ANTIGEN
15	YP_001387133.1	<b>0.4053</b>	Probable ANTIGEN
16	YP_001387258.1	0.3751	<b>Probable NON-ANTIGEN</b>
17	YP_001387314.1	<b>0.5220</b>	Probable ANTIGEN
18	YP_001386372.1	<b>0.4096</b>	Probable ANTIGEN
19	YP_001387500.1	0.3101	<b>Probable NON-ANTIGEN</b>
20	YP_001388739.1	<b>0.6415</b>	Probable ANTIGEN
21	YP_001388796.1	<b>0.7614</b>	Probable ANTIGEN
22	YP_001389355.1	<b>0.6086</b>	Probable ANTIGEN

**Bcpred result based on Vaxijen and TMHMM Scores:** The Bcpred results recommended that predicted peptides among 22 proteins only 19 are having Bcpred score >0.8 shown (**Table 2**). These 19 proteins were exposed to TMHMM again to identify exo-membrane constituencies and from them 15 proteins were concluded with Bcpred score >0.8 and exo-membrane topology. The

Superficial exposed B-cell epitope peptides of 15 finalized proteins were analyzed using Vaxijen (threshold=0.4) to check the antigenicity and all peptides except one are having Vaxijen score >0.4 results are précised in Table 3. So 15 membrane proteins, among 22 were finalized as antigenic exo-membrane B-epitope targets shown in (**Table 3**).

<b>Table 2: TMHMM result of 17 finalized proteins from VexiJen score</b>			
<b>S.No.</b>	<b>PROTEIN</b>	<b>TMHMM RESULTS</b>	<b>LENGTH OF PROETIN</b>
<b>1</b>	YP_001386116.1	1- exo peptide	1-133
<b>2</b>	YP_001387255.1	2- exo peptide	1-90 209-572
<b>3</b>	YP_001386503.1	5- exo peptide	46- 59 117 - 167 231 - 239 298- 349 400 - 433
<b>4</b>	YP_001386587.1	5- exo peptide	1 - 111 164 - 196 262- 275 342 - 350 409 - 422
<b>5</b>	YP_001386886.1	3- exo peptide	30 - 55 208 - 232 110 - 123
<b>6</b>	YP_001387027.1	4-exo peptide	30 - 67 125 - 128 200 - 203 253 - 268
<b>7</b>	YP_001387384.1	2-exo peptide	30 - 43 102 - 190
<b>8</b>	YP_001387457.1	1-exo peptide	1-212
<b>9</b>	YP_001387707.1	2- exo peptide	186 - 211 393- 396
<b>10</b>	YP_001388265.1	1-exo peptide	29 - 556
<b>11</b>	YP_001387020.1	5-exo peptide	1 - 14 106 - 147 203 - 211 323 - 352 405 - 408
<b>12</b>	YP_001387133.1	3-exo peptide	36 - 97 156 - 174 259 - 277
<b>13</b>	YP_001387314.1	3-exo peptide	1 - 19 77 - 90 157 - 188
<b>14</b>	YP_001386372.1	7-exo peptide	28 - 41 87 - 109 176 - 189 256 - 302 381 - 394 461 - 474 544 - 546
<b>15</b>	YP_001388739.1	1-exo peptide	1 - 3
<b>16</b>	YP_001388796.1	1-exo peptide	1 - 3
<b>17</b>	YP_001389355.1	4-exo peptide	46 - 64 175- 216 267 - 270 325 327

**Table 3: Summarized results of BCPred, TMHMM and VexiJen of finalized 15 proteins**

S.No.	Protein name	Total no of epitope	Amino acids position	Bcpred epitope sequence	Bcpred score	Vaxijen score
1	YP_001386116.1	1	41	NMITYLKPTITKYIDLNGNK	0.845	0.6981
2	YP_001387255	4	306	SMEETGASAEEMSATSEEIE	0.973	1.3210
			119	EKNLNEFRQRREEVFKVAKE	0.925	0.1372
			22	IIVGITGYFNAKSNAIKK	0.901	0.6987
			370	IDSATKDAIEKSKTIEEINV VTTNIISKSIINPLKESVEY	0.82 0.872	0.6195
3	YP_001386503	2	111	IGTLINPTKGVDPSTMQKI	0.906	<b>0.4247</b>
			400	DAPATLLNSTGNTVCAMMIT	0.851	<b>0.5935</b>
4	YP_001386587	3	32	FNIKSEEKANDEEVKSIKGV	0.985	<b>1.5689</b>
			445	GFDDPVEQNKEEQKKVMEEN	0.98	<b>0.7432</b>
			84	SSKDNNEKKKEGIVDRVLSV	0.957	<b>1.2129</b>
5	YP_001386886	1	211	TRMKTEIPTSIFLNMSTGD	0.879	<b>0.4487</b>
6	YP_001387027	1	217	TYPIVMFPFIQSGDRTIASS	0.873	<b>1.2066</b>
7	YP_001387384	1	162	TLMVAGNIPGKTQTIPTAIY	0.761	<b>0.5783</b>
8	YP_001387457	3	18	IEIFKTININTRELEDKGRK	0.963	<b>0.8082</b>
			103	LPSFKLDQGYNRKKIATKYP	0.873	<b>1.0699</b>
			55	KAKDVETYVEYGAADIGIVG	0.83	<b>0.4053</b>
9	YP_001387707	3	41	NITNKNIFKIRSEKITLKDC	0.825	<b>0.9307</b>
			296	FKENIKNLNKNIKDGKSISI	0.823	<b>0.7517</b>
			193	FFYKRYKIKYAVDNLKIKTP	0.814	<b>0.7133</b>
10	YP_001388265	2	117	KSGKDIKNTCTTLGANRFQT	0.998	<b>0.9145</b>
			51	KKIMIQINHTLMSIPPVLMG	0.893	<b>0.7043</b>
11	YP_001387020	10	79	VPSAFKDNKEEKDEEKLLTI	0.996	<b>0.7198</b>
			524	VVFIPKIKTKGESAGTTSVP	0.996	<b>1.1954</b>
			192	NRVKDNKTSKIIYKDLQGE	0.992	<b>1.2277</b>
			106	NEKYDITQNQTIDSSGKNYY	0.978	<b>0.9687</b>
			213	ISKEAEEPKNINPKLTLDS	0.962	<b>1.3843</b>
			128	DKITYEKLKDIKGVKGFYTY	0.953	<b>0.6478</b>

			313	SFKCTGEFESNKKGTHGSFS	0.928	<b>0.8250</b>
			376	EQQGAIQMEKGEKPNLSDGT	0.909	<b>0.7079</b>
			487	KTGTNERQEVNDKKGMEHLS	0.905	<b>1.3061</b>
			149	KQEVNKNINDKKEAWKLENM	0.881	<b>0.8601</b>
<b>12</b>	YP_001387133	3	223	DKIIKEPAGGAHKNLNKMAE	0.853	<b>0.5951</b>
			25	ARLKERPTALDYINIIFDDF	0.823	<b>0.5965</b>
			119	DTQGAFCGIDAEERGQGEA	0.782	<b>1.0672</b>
<b>13</b>	YP_001387314	5	84	IIGVPVAIAFGGPGAVFWMW	1	<b>0.9976</b>
			242	IFKGAFTPQAAVGGFGGATL	0.994	<b>0.4582</b>
			266	RWGTARGTYSNEAGMGSAPI	0.95	<b>0.6914</b>
			48	KETFGKMFSPAEGETTTP	0.933	<b>0.6455</b>
			152	MCSFCFMIEIIPSISTQSL	0.768	<b>1.0821</b>
<b>14</b>	YP_001388739	1	43	KPNKIHIMGTDSMGRDVFSR	0.802	<b>0.6468</b>
<b>15</b>	YP_001388796	3	92	DTDKKEDMDKKEEKSAKEK	0.999	<b>1.7980</b>
			24	YVIIQTGPDRVQVQISKSGI	0.994	<b>0.5201</b>
			64	PEKKRKKKYNEMLNSISVND	0.921	<b>0.9828</b>

**B –cell derived T-cell:** Each selected B-cell epitope was examined for identification of T-cell within the B-cell epitope sequence. For the first level screening, Propred-I (47 MHC Class-I alleles), Propred (51 MHC Class-II alleles), and Mhcpred (DRB1\*0101 allele) were used to isolate common T-cell epitope that share B-cell epitope sequence, can interact with both the MHC classes with highest number, and specifically interact with DRB1\*0101 (as the DRB1\*0101 is commonest bound allele, therefore the interaction epitope produce better antigenic response).

At the second level of screening, identified peptides in the first screen were used to predict their binding abilities to >1000 MHC alleles using T-Epitope Designer and that bind to >100% alleles were selected. Similarly, as **A\*0201**, **A\*0204**, and **B\*2705** alleles are mostly used in various prediction methods, we set the cut off that selected

peptides must bind to these three HLA molecules and T-epitope Designer was also used for this purpose. Then selected epitope % work done at T-epitope designer tool (**Table 5**). Since the frequency of **DRB1\*0101** and **DRB1\*0401** alleles of MHC Class-II is 20-50% in all. We selected T-epitope that interacted with these two HLA molecules using Mhcpred as described in methods. The final list of epitope was made with non-overlapping peptide sequences that confirm the above mentioned criteria and Vaxijen and IC50 scores. with highest antigenicity and those bind more than 15 MHC molecules comprising of both the MHC class I and II alleles and less than 100 nM IC50 scores for DRB1\*0101 were selected. From the result 4 proteins with 5 peptides among 15 were fulfilled the criteria and predicted as T-epitope candidates are shown in (**Table 4 as \***).

**Table 4: Common epitopes from each protein that can produce both the B- and T-cell mediated immunity are represents along with their various parameters. Epitopes selected as vaccine candidates are labeled as \***

S.No.	Protein id	Bepred predicted epitope sequence	Number of MHC class I binding alleles PropredI	Number of MHC class II binding alleles Propred	MhcPred predicted epitope sequence	Vexijen score based on MhcPred sequence	IC50 value of epitopes for DRB1*0101 (MHCPRD)	Total number of MHC binding alleles
1	YP_001386111	NMITYLKPTITKYIDLNGNK	0	23	YLKPTITKY TITKYIDLN NMITYLKPT KYIDLNGNK MITYLKPTI ITKYIDLNG	0.7397 0.6176 0.1075 1.5123 0.2926 1.0295	0.24 30.90 34.12 34.83 40.36 53.58	23
2	YP_001387255	SMEETGASAEEMSATSEEIE	0	0	MEETGASAE EMSATSEEI EETGASAEI SMEETGASA EEMSATSEE ETGASAEEM SAEEMSATS AEEMSATSE GASAEEMSA	2.0598 0.9682 1.7880 1.3794 1.5282 1.3154 1.2224 1.2644 1.0501	13.52 29.85 30.20 37.58 38.99 55.85 57.15 61.80 68.71	0
		EKNLNEFRQRREEVFKVAKE	0	1	EEVFKVAKE NLNEFRQR FRQRREEVF QRREEVFKV KNLNEFRQR NEFRORREE LNEFRORRE REEVFKVAK EFRORREEV	0.8125 0.4323 0.1051 0.0243 0.7049 1.5227 1.1472 0.8793 0.3925	13.80 15.74 25.82 28.71 29.38 39.17 43.25 89.54 96.61	1
		IIVGITGYFNAKSNKAIKK	0	25	YFNAKSNKA YYFNAKSNK IIVGITGY GYYFNAKSN NAKSNKAIK GITGYFNA	1.7583 1.4327 0.3207 1.0890 1.2196 0.9462	0.82 4.37 12.39 33.19 54.95 75.34	25
		VTTNIISKSIINPLKESVEY	0	7	ISKSIINPL SIINPLKES IINPLKESV IISKSIINP INPLKESVE VTTNIISKS NIISKSIIN KSIINPLKE	-0.8236 -1.0420 0.8115 -0.6858 -0.3249 -0.2138 -0.3920 -1.5567	1.83 9.73 13.90 17.02 19.32 28.71 52.48 69.02	7
		IDSATKDAIEKSKTIEEINV	0	1	IDSATKDAI IEKSKTIEE SATKDAIEK ATKDAIEKS	0.9744 0.5307 0.3668 0.3223	9.12 22.86 26.67 37.33	1
		3	YP_001386503	IGTLIINPTKGVDPSTMQKI	0	1	IGTLIINPT IINPTKGVDP INPTKGVDP KGVDPSTMQ GVDPSTMQK NPTKGVDP	-0.8689 1.3924 1.3938 1.6148 0.7043 1.4531
DAPATLLNSTGNTVCAMMIT	0			8	DAPATLLNS GNTVCAMMI NSTGNTVCA STGNTVCAM	0.3571 1.0599 1.0920 0.4179	1.05 8.63 30.20 62.37	8
4	YP_001386587	FNIKSEEKANDEEVKSIKGV	5	1	KSEEKANDE KANDEEVKS NIKSEKAN EEKANDEEV	1.8137 2.2451 0.9825 1.8589	3.26 19.36 35.97 56.62	6
			0	0	ENKSEEKA IKSEEKAND DDPVEQNKE QNKEEQKKV FDDPVEQNK KEEQKKVME EEQKKVMEE GFDDPVEQN	1.4696 1.8361 0.2730 1.8256 0.6671 0.4609 0.2955 0.4641	81.66 86.30 11.32 16.48 16.94 50.47 95.72 96.38	0
		SSKDNNEKKKEGIVDRVLSV	0	0	GIVDRVLSV EGIVDRVLS	-0.1952 0.4301	9.44 23.17	0
5	YP_00138686	TRMKTEIPTSIFLNMSTGD	0	4	IPTSIFLNM EIIPTSIFL PTSIFLNM IPTSIFLN RMKTEIPT IFLNMSTGD	0.1837 0.7217 -0.2156 0.2835 0.0357 0.3659	6.19 7.55 7.71 12.33 16.60 73.96	4
6	YP_0013870.1	TYPIVMFPIQSGDRTIASS	0	45	SGDRTIASS IVMFPIQS PIVMFPIQ FIQSGDRTI MFPFIQSGD YPIVMFPI	0.7581 1.2194 2.2772 0.3658 0.7988 3.3778	4.55 12.33 16.87 22.70 38.02 40.93	



					QSGDRTIAS IQSGDRTIA TYPIVMFPF	0.9732 1.2340 3.1066	58.88 60.81 65.16	
7	YP_001387384	TLMVAGNIPGKTQTIPTAIY	0	14	KTQTIPTAI TLMVAGNIP PGKTQTIPT TQTIPTAIY NIPGKTQTI	0.4205 0.0788 0.5960 0.5508 0.2571	26.98 29.92 37.76 47.86 61.94	14
8	YP_001387457*	IEIFKTININTRELEDKGRK*	6	38	NINTRELED IEIFKTINI EIFKTININ INTRELEDK IFKTININT ININTRELE FKTININTR TININTREL NNTRELEDKG	1.1550 -0.3309 0.4764 1.4586 1.0627 2.3701 1.3949 1.4760 0.8280	10.19 27.73 30.13 31.48 34.43 35.48 37.67 72.28 93.76	44
		LPSFKLDQGYNRKKIATKYP	0	17	NRKKIATKY SFKLDQGYN FKLDQGYNR GYNRKKIAT YNRKKIATK PSFKLDQGY	0.6706 0.5881 0.7713 -0.4442 0.6158 0.7628	4.32 26.73 47.53 64.71 68.55 85.31	17
		KAKDVETVVEYGAADIGIVG*	8	8	YVEYGAADI TYVEYGAAD KDVETVVEY KAKDVETVY EYGAADIGI YGAADIGIV	0.1827 0.2998 0.2379 0.7194 0.6886 0.8185	5.86 31.70 42.17 43.95 59.16 97.72	16
9	YP_001387707*	NITNKNIFKIRSEKITLKDC	0	24	SEKITLKDC IRSEKITLK KNIFKIRSE IFKIRSEKI NITNKNIFK NIFKIRSEK TNKNIFKIR RSEKITLKD KIRSEKITL ITNKNIFKI	1.1059 1.5512 0.7365 1.8222 0.2600 0.7830 0.9905 1.1855 0.9130 0.5455	22.18 28.05 29.04 38.02 41.98 42.17 53.09 55.98 80.35 89.74	24
		FKENIKNLNKNIKDGKSISI*	3	27	NIKDGKSIS NIKNLNKNI FKENIKNLN ENIKNLNKN IKDGKSISI	0.8192 -0.5160 0.4539 0.1772 2.1246	6.61 9.29 25.41 50.35 98.40	30
		FFYKRYKIKYAVDNLKIKTP	0	23	FYKRYKIKY KIKYAVDNL YAVDNLKIK RYKIKYAVD KYAVDNLKI YKRYKIKYA AVDNLKIKT	0.8634 -0.2952 2.1610 1.0271 0.9084 1.1921 1.8388	1.02 11.40 16.37 30.62 35.73 35.73 54.33	0
10	YP_001388265	KSGKDIKNTCTTLGANRFQT	0	8	CTTLGANRF TCTTLGANR KNTCTTLGA DIKNTCTTL NTCTTLGAN SGKDIKNTC TLGANRFQT	0.3731 0.6461 0.5892 0.6486 1.0142 1.0292 0.4561	3.49 6.49 8.22 16.87 21.13 32.43 79.07	0
		KKIMIQINHTLMSIPPVLMG	0	50	QINHTLMSI INHTLMSIP KIMIQINHT MIQINHTLM TLMSIPPVL HTLMSIPPV NHTLMSIPP IQINHTLMS MSIPPVLMG	0.1541 -0.1214 0.9198 1.0504 1.4452 0.6236 0.3938 0.7890 0.7154	3.79 10.35 15.35 25.35 26.18 29.51 30.27 37.41 68.55	50
11	YP_001387020*	VPSAFKDNKEEKDEEKLLTI	0	0	KDEEKLLTI EEKDEEKLL PSAFKDNKE DNKEEKDEE FKDNKEEKD EKDEEKLLT	-0.5857 0.4140 -0.2012 2.2560 2.4049 -0.1784	18.62 31.70 33.96 35.56 45.19 89.95	0
		VVFIPKIKTKGESAGTTSVP*	2	42	KIKTKGESA FIPKIKTKG ESAGTTSVP	1.4006 0.0516 0.9316	15.63 35.16 76.03	44
		NRVKDNKTSKIIYKKDLQGE	0	3	IYKKDLQGE IIYKKDLQG KIIYKKDLQ DNKTSKIIY RVKDNKTSK NKTSKIIYK	1.5027 1.2054 0.1733 -0.2053 2.0850 0.0903	14.39 25.82 26.67 30.69 37.33 69.82	3
		NEKYDITQNQTIDSSGKNYY	0	13	KYDITQNQT IDSSGKNYY TIDSSGKNY TQNQTIDSS DITQNQTID	0.7658 1.8068 0.9657 0.8574 0.5253	2.88 16.60 18.28 19.05 24.10	13

					QNQTIDSSG ITQNQTIDS NEKYDITQN EKYDITQNO NQTIDSSGK YDITQNQTI	1.3929 01.220 0.8988 1.1749 1.1139 0.5493	39.26 39.36 45.92 76.91 79.25 90.99	
		ISKEAEPPKSNINPKLTLDS	0	32	NINPKLTL EAEPPKSN INPKLTLDS EPKSNINPK ISKEAEPPK AEEPKNIN	2.3057 0.4077 1.7361 1.3404 1.4756 0.7647	2.96 14.59 17.66 21.38 86.10 87.90	32
		DKITYEKLKDIKGVKGFYTY	0	4	KITYEKLK YEKLKDIK ITYEKLKDI KLKDIKGVK DIKGVKGFY TYEKLKDIK	0.5463 0.8069 -0.0591 1.0099 0.2432 0.2710	7.21 12.30 16.11 21.68 40.18 88.31	4
		SFKCTGEFESNKKGTHGSFS	0	6	FKCTGEFES TGEFESNKK FESNKKGTH SFKCTGEFE	0.1155 0.1017 2.6654 -0.1207	54.45 55.72 59.57 76.38	6
		EQQGAIQMEKGEKPNLSDGT	0	0	GEKPNLSDG MEKGEKPNL EQQGAIQME EKGEKPNLS IQMEKGEKP QQGAIQMEK	0.5964 1.4655 0.6455 1.5979 0.4918 0.4918	39.99 44.67 55.21 61.38 67.14 83.56	0
		KTGTNERQEVNDKGMKMEHLS	0	7	QEVNDKGMK TGTNERQEV TNERQEVND NERQEVNDK NDKGMKMEHL EVNDKGMKME	0.6192 1.5187 0.9566 1.5214 0.6646 0.6720	19.91 37.33 44.87 46.24 50.93 98.40	7
		KQEVNDKNINDKKEAWKLENM	0	0	NDKKEAWKL NINDKKEAW EVDKNINDK QEVNDKNIND INDKKEAWK KQEVNDKNIN KNINDKKEA KEAWKLENM	0.6709 -0.6496 1.5209 0.8674 0.3138 1.0295 1.2620 0.3980	8.20 20.37 30.97 33.57 55.59 56.36 60.67 70.79	0
12	YP_001387133	DKIIEPAGGAHKNLNKMAE	0	0	IIEPAGGA GAHKNLNKM	0.9651 0.4107	3.65 23.01	1
		ARLKERPTALDYINIIFDDF	0	3	KERPTALDY YINIIFDDF DYINIIFDD TALDYINII ERPTALDYI RLKERPTAL RPTALDYIN	0.1009 1.3204 0.7424 -0.1944 0.2703 0.3706 -0.0005	0.75 10.23 11.38 12.56 13.61 20.28 74.13	3
		DTQGAFCGIDAERGGQGEA	0	2	FCGIDAER GIDAERGGQ	1.0394 1.1701	26.36 41.88	2
13	YP_001387314*	IIGVPVAIAFGGPGAVFWMW	0	18	IGVPVAIAF FGGPGAVFW IIGVPVAIA	1.0727 1.0641 0.7140	18.54 20.04 27.73	18
		IFKGAFTPQAAVGGFGGATL*	4	15	IFKGAFTPQ AFTPQAAVG GAFTPQA FTPQA FKGAFTPQA	0.0284 1.0686 0.8912 1.4438 0.0834	12.91 13.68 35.32 42.56 69.02	19
		RWGTARGTYSNEAGMGSAPI	0	2	TYSNEAGMG WGTARGTYS YSNEAGMGS EAGMGSAPI TARGTYSNE ARGTYSNEA	1.3142 1.3114 1.8533 1.1848 1.1406	22.49 31.92 61.52 62.09 62.95 93.33	2
		KETFGKMFSPKPAEGEGTITP	0	10	FGKMFSPKA FSKPAEGEG KETFGKMFS TFGKMFSKP	0.2997 2.2657 -0.2576 -0.5614	4.98 30.20 43.25 73.11	10
		MCSFCFMIHPSISTQSLS	0	34	PSISTQSLS IIPSIQS FMIEHPSI CFMIEHPS EIIPSIQS MIEHPSIS FCFMIEIIP SFCFMIEII	1.1298 0.7081 0.7752 1.4870 0.5050 0.5922 1.7171 1.8738	3.66 5.65 10.59 16.83 20.99 24.55 30.55 30.55	34
14	YP_001388739	KPNKIHMGTDSMGRDVFSTR	0	31	HIMGTDSMG MGTDSMGRD DSMGRDVF KIHMGTDS TDSMGRDVF SMGRDVFSTR IMGTDSMGR	1.1178 2.0171 0.2259 1.1923 0.8134 0.1891 1.3634	23.33 50.82 56.89 57.68 64.57 84.53 95.50	31

15	YP_001388796	DTDKKEDMDKKEEKSAKEK	0	0	KEDMDKKEE DMDKKEEK KKEEKSAKE EDMDKKEEK	1.7079 1.7928 2.2276 2.0821	38.11 65.77 68.08 89.95	0
		YVIIQTGPDRVKVQISKSGI	0	28	RVKVKQISKS IIQTGPDRV PDRVVKVQIS YVIIQTGPD QTGPDRVKV IQTGPDRVK KVQISKSGI	0.7288 1.1728 0.8000 0.3088 0.9551 1.6724 0.1868	2.07 12.85 20.61 26.24 29.58 50.93 71.94	28
		PEKKRKKKYNEMLNSISVND	0	8	KYNEMLNSI YNEMLNSIS MLNSISVND KKYNEMLNS PEKKRKKKY EMLNSISVN KRKKKYNE NEMLNSISV	0.1323 0.2446 1.0659 0.4477 1.6571 1.1054 0.7381 0.1941	3.68 9.25 28.12 34.28 39.54 46.34 55.21 58.21	8

Table 5: Final selection of T-epitopes from 22 proteins *Clostridium botulinum*

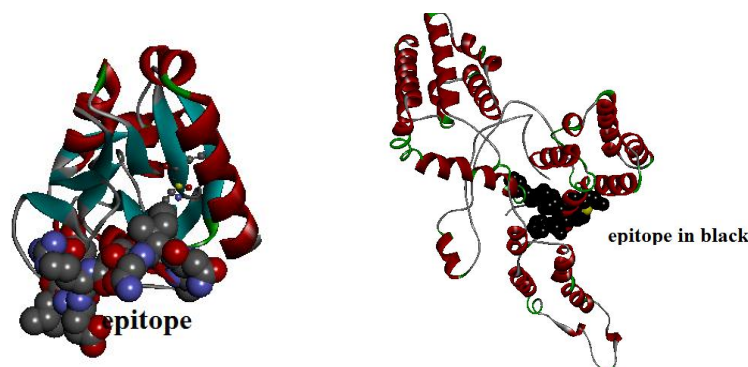
S.No.	Protein	Bcpred predicted epitope sequence	Epitopes	T-Epitope Designer A*0201	T-Epitope Designer A*0204	T-Epitope Designer B*2705	Mhcpred (IC50 value) DRBI*0101	MHCPred (IC50 values) DRBI*0401
1	YP_001387457	IEIFKTININTRELEDKGRK	NINTRELED	414.90	-238.08	50.09	10.19	774.46
			EIFKTININ	-666.14	-581.60	268.50	30.13	505.82
			INTRELEDK	387.71	-3.97	2050.80	31.48	1778.28
			IFKTININT	692.05	698.62	-109.28	34.43	2233.57
			ININTRELE*	564.12	160.73	1472.59	35.48	1169.50
2	KAKDVETVVEYGAADIGIVG	FKTININTR	116.12	-404.02	1737.06	37.67	246.60	
		TININTREL	288.37	-282.39	575.60	72.28	458.14	
		KAKDVETVY	476.60	-0.30	-194.58	43.95	1545.25	
		EYGAADIGI	-1176.99	-832.31	-1901.75	59.16	195.88	
		YGAADIGIV	-1421.67	-1347.90	-555.62	97.72	405.51	
3	YP_001387707	FKENIKNLNKNIKDGKSISI	NIKDGKSIS	-422.70	-1335.83	-405.58	6.61	1256.03
			FKENIKNLN*	680.97	152.62	1577.69	25.41	228.56
			IKDGKSISI	44.32	-36.26	-200.68	98.40	1479.11
4	YP_001387020	VVFIPKIKTKGESAGTTSVP	KIKTKGESA	302.16	-379.47	994.38	15.63	993.12
			ESAGTTSVP	-1076.85	-1386.53	-117.11	76.03	258.82
5	YP_001387314	IFKGAFTPQAAVGGFGGATL	IFKGAFTPQ	-9.95	-129.75	364.70	12.91	788.86
			AFTPQAAVG	-826.56	-926.19	-267.74	13.68	---
			GAFTPQAAV	-203.67	-108.78	-865.36	35.32	797.99
			FTPQAAVGG	-417.68	-450.27	681.70	42.56	193.20
			FKGAFTPQA	-484.89	-403.49	768.31	69.02	96.38

**Clusters and folding:** Homology modeling for each full length protein was carried out using Phyre version 2.0 Web-server and best models were selected based on super families and E-values of templates. The 3D folding and clusters of in folded protein were analyzed to confirm the exo-membrane topology of this using Pepitope server. Pepitope was fed with Phyre derived 3D structure of each protein and

all identified from the same protein to analyze the linear alignment of on the corresponding protein and to determine the epitope clusters and exo-membrane position of in the folded proteins shown in (Figure 1). And best clusters calculated by Pepitope were having scores 45.551 residues no 12 for peptide **ININTRELE** and 10.122 residues no 9 for peptide **FKENIKNLN** (Table 6).

Table 6: Screening of T-epitopes using T-epitope Designer

Protein names	Epitope	T-Epitope Designer (no of binders with HLA molecules)	comments	lowest score	highest score
YP_001387457	<a href="#">ININTRELE</a>	100%	almost all values are positive .mostly bind with A,B,C alleles.	24.09(A) 82.35(B)	2025.15(B*1561) 1635.00(A*3401)
YP_001387707	<a href="#">FKENIKNLN</a>	90%	mostly positive for all B and C alleles	1.82(A-alleles) 87.07(B alleles )	1100.77(A alleles) 1848.06(B alleles)



**Fig. 1: Fold level characterization of cluster and topology of best using Pepitope (in red and black)**

### CONCLUSION

In this study by using computational approaches based on sequence structure, QSAR, simulation, and fold level analysis, we identified two potential T- derives from antigenic B-cells epitope of twenty two exo-membrane essential proteins of *C. botulinum*. Selected T- ["INTRELEDK" from YP\_001387457, "FKENITKNL" from YP\_001387707] are antigenic and have much potential to interact most common human HLA alleles (A\*0201, A\*0204, B\*2705, DRB1\*0101, and DRB1\*0401). These are also found to interact with >75% of HLA molecules in a binding screening using T-Epitope Designer (that contains >1000 HLA molecules). Therefore these selected are likely to induce both the B-cell and T-cell mediated immune responses. Homology and simulation results also support the suitability of these as vaccine candidates. However, there are several pitfalls in developing a good vaccine and moreover there is lack of proper experimental disease model for botulinum suitable animal model should be used for experimental validation of these to confirm these "Insilco" results.

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